

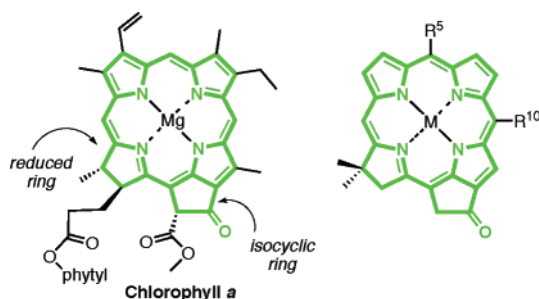
A New Route for Installing the Isocyclic Ring on Chlorins Yielding 13¹-Oxophorbines

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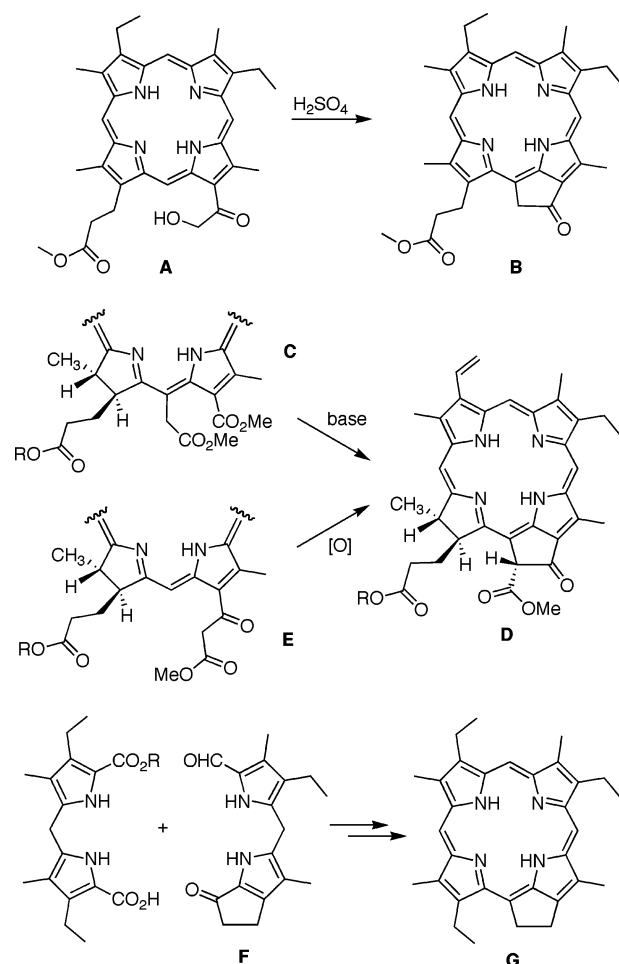
A new route to 13¹-oxophorbines, the parent macrocycle of chlorophylls, begins with the synthesis of a 13-bromochlorin. Pd-mediated coupling of the latter with tributyl(1-ethoxyvinyl)tin and subsequent acidic hydrolysis afforded the 13-acetylchlorin (**1**). Treatment of **1** with NBS afforded the 15-bromo analogue in 70% yield. Pd-mediated α -arylation closed the isocyclic ring to give the 13¹-oxophorbine (**2**) in 85% yield. Facile access to 13¹-oxophorbines should enable a variety of spectroscopic studies and diverse applications.

Introduction

The fundamental skeleton of chlorophylls is a phorbine, which differs from porphyrins in containing one reduced pyrrole ring and a five-membered exocyclic ring. The exocyclic ring (known as the isocyclic ring) is ortho-perifused across the 13- and 15-positions and contains a 13¹-oxo group.¹ Chlorophyll *a* absorbs strongly in the blue (B band, ~430 nm) and the red (Q_y band, ~660 nm) regions of the visible spectrum. The 13-keto group, which is conjugated with the π -electrons of the macrocycle, causes a significant red-shift and intensification of the Q_y band compared to synthetic chlorins lacking a 13-keto substituent.² Intense absorption in the red region is attractive for applications encompassing solar cells,³ medical imaging,⁴ and photodynamic therapy.⁵ Moreover, the 13-keto substituent is an essential motif for self-assembly of certain chlorophyll species.⁶

Surprisingly few routes are known for the construction of the isocyclic ring (Scheme 1).⁷ Fischer reported the dehydration of a (hydroxymethylcarbonyl)porphyrin using concentrated H₂-

SCHEME 1



SO₄ to give the “pheoporphyrin” bearing the isocyclic ring (**A** → **B**),⁸ and Dieckmann cyclization to convert chlorin e₆ trimethyl ester to methyl pheophorbide *a* (**C** → **D**).^{9,10} The Dieckmann cyclization has been carried out using KOH/pyridine,⁹ sodium methoxide in methanol/acetone,¹⁰ potassium *tert*-butoxide/pyridine,¹¹ sodium bis(trimethylsilylamide),¹² or potassium *tert*-butoxide/collidine.¹³ Kenner employed oxidative cyclization with a porphyrin bearing a β -ketoester to give the pheoporphyrin,¹⁴ a route extended by Smith for conversion of a chlorin to the methyl pheophorbide *a* (**E** → **D**).¹⁵ Finally, a

(6) Balaban, T. S.; Tamiaki, H.; Holzwarth, A. R. *Top. Curr. Chem.* **2005**, 258, 1–38.

(7) Pavlov, V. Y.; Ponomarev, G. V. *Chem. Heterocycl. Compd.* **2004**, 40, 393–425.

(8) Fischer, H.; Laubereau, O. *Justus Liebigs Ann. Chem.* **1938**, 535, 17–37.

(9) Fischer, H.; Lautsch, W. *Justus Liebigs Ann. Chem.* **1937**, 528, 265–275.

(10) Fischer, H.; Oestreicher, A. *Justus Liebigs Ann. Chem.* **1940**, 546, 49–57.

(11) (a) Smith, K. M.; Bisset, G. M. F.; Bushell, M. J. *Bioorg. Chem.* **1980**, 9, 1–26. (b) Smith, K. M.; Bushell, M. J.; Rimmer, J.; Unsworth, J. F. *J. Am. Chem. Soc.* **1980**, 102, 2437–2448. (c) Smith, K. M.; Bisset, G. M. F.; Bushell, M. J. *J. Org. Chem.* **1980**, 45, 2218–2224.

(12) Gerlach, B.; Brantley, S. E.; Smith, K. M. *J. Org. Chem.* **1998**, 63, 2314–2320.

(13) Pallenberg, A. J.; Dobhal, M. P.; Pandey, R. K. *Org. Process Res. Dev.* **2004**, 8, 287–290.

[†] Equal contributions by both authors.

(1) Scheer, H. In *Chlorophylls*; Scheer, H., Ed.; CRC Press: Boca Raton, FL, 1991; pp 3–30.

(2) Boldt, N. J.; Donohoe, R. J.; Birge, R. R.; Bocian, D. F. *J. Am. Chem. Soc.* **1987**, 109, 2284–2298.

(3) Linke-Schaetzel, M.; Bhise, A. D.; Gliemann, H.; Koch, T.; Schimmel, T.; Balaban, T. S. *Thin Solid Films* **2004**, 451, 16–21.

(4) Licha, K. *Top. Curr. Chem.* **2002**, 222, 1–29.

(5) Nyman, E. S.; Hynninen, P. H. *J. Photochem. Photobiol. B: Biol.* **2004**, 73, 1–28.

TABLE 1. Spectral Properties of Chlorins and 13¹-Oxophorbines^a

compound	λ_B (fwhm) in nm	$\log \epsilon_B$	λ_{Q_y} (fwhm) in nm	$\log \epsilon_{Q_y}$	$\Delta\nu_{Q_y}$ (cm ⁻¹) ^b	I_B/I_{Q_y} ^c	λ_{em} (nm) ^d	$\Delta\nu$ (cm ⁻¹) ^e	Φ_f ^f
Zn-8	412 (13) ^g	5.27	608 (11)	4.64	0	4.9	611	80	0.062
Zn-1	424 (17) ^h	5.30	635 (14)	4.85	700	2.8	639	100	0.25
Zn-2	431 (16) ⁱ	5.15	643 (11)	4.84	900	2.1	646	70	0.28
8	414 (34) ^j	4.95	641 (9)	4.45	0	3.3	643	50	0.23
1	423 (36)	5.14	661 (13)	4.72	470	2.6	665	90	0.25
2	417 (49) ^k	5.00	660 (11)	4.71	450	2.0	663	70	0.37
Zn-Pheo a^{l,m}	423 (38)	5.09	653 (18)	4.96	—	1.4	657	90 ⁿ	0.23 ⁿ
Pheo a^{o,p}	410 (63) ^q	4.97	667 (21) ^q	4.65	—	2.1	675 ^q	180 ^r	0.28 ^r
Chl a^{s,t}	433 (39) ^q	5.01	666 (19) ^q	4.89	—	1.3	671 ^q	110 ^q	0.325 ^q

^a In toluene at room temperature unless noted otherwise. ^b The redshift of the Q_y band relative to that of the parent chlorin (**8** or **Zn-8**). ^c Ratio of the intensities of the B and Q_y bands. ^d Excitation was performed at the λ_{max} of the B band. ^e Stokes shift. ^f Determined with λ_{exc} at the B band maximum using chlorophyll *a* as a standard ($\Phi_f = 0.322$) unless noted otherwise (see Supporting Information). ^g Shoulder at 391 nm. ^h Shoulder at 402 nm. ⁱ Shoulder at 409 nm ($\log \epsilon = 4.73$). ^j Shoulder at 398 nm. ^k Shoulder at 431 nm ($\log \epsilon = 4.97$). ^l In diethyl ether. ^m Absorption data from ref 30. The absorption spectrum is essentially identical in $CHCl_3$.²⁹ ⁿ Reference 32. The Φ_f value was calculated using data from ref 32 with chlorophyll *a* as standard. A value of $\Phi_f = 0.17$ in diethyl ether/petroleum ether has been reported.³¹ ^o In ethanol. ^p Absorption and emission data from ref 33. ^q This work. ^r In ethanol. The Φ_f in toluene was found to be as follows: Pheophorbide *a* (0.26), pyropheophorbide *a* (0.29), pyropheophorbide *a* methyl ester (0.30). ^s In benzene. ^t Absorption data from ref 28. ^u Reference 27.

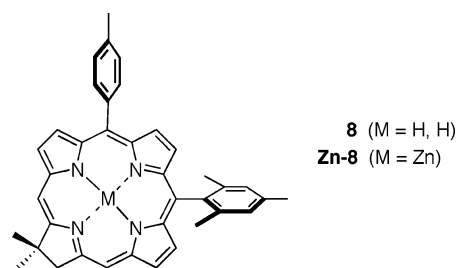
and tributyl(1-ethoxyvinyl)tin (40 mM) in the presence of 15 mol % of $(PPh_3)_2PdCl_2$ in THF for 20 h followed by hydrolysis with 10% aqueous HCl gave 13-acetylchlorin **1** in 74% yield. A streamlined procedure including demetalation, Pd-coupling, and acidic workup gave **1** in 68% yield starting from **Zn-6**. The free-base 13-acetylchlorin **1** was metalated with $Zn(OAc)_2 \cdot 2H_2O$ or $Cu(OAc)_2 \cdot H_2O$ to obtain **Zn-1** or **Cu-1**, respectively. The chlorins were characterized by mass spectrometry (LD-MS, FAB-MS) and absorption, fluorescence, and ¹H NMR spectroscopy. The X-ray structure of **Cu-1** confirmed the presence of the acetyl group at the 13-position of the chlorin macrocycle (Supporting Information).

C. Isocyclic Ring Installation. The α -arylation of aliphatic ketones has been carried out on a wide variety of aryl substrates, including aryl bromides under Pd-mediated coupling conditions.²⁵ Treatment of **1** to conditions for 15-bromination (1 equiv of NBS at room temperature for 1 h)²⁶ gave the 15-bromochlorin **7** in 70% yield. The reaction of **7** under conditions for α -arylation [$(PPh_3)_2PdCl_2$ in the presence of Cs_2CO_3 in toluene at reflux]²⁵ for 24 h gave oxophorbine **2** in 85% yield. A combined procedure of bromination and Pd-mediated ring closure gave oxophorbine **2** in 44% yield from **1**. Treatment of **2** with $Zn(OAc)_2 \cdot 2H_2O$ at room temperature gave **Zn-2** in 85% yield. Oxophorbines **2** and **Zn-2** were characterized by mass spectrometry (LD-MS, FAB-MS) and absorption, fluorescence, IR and ¹H NMR spectroscopy.

D. Spectral Properties. The spectral properties of the 13-acetylchlorins and 13¹-oxophorbines are listed in Table 1, accompanied by those of chlorophyll *a* (**Chl a**),^{27,28} the zinc (**Zn-Pheo a**)^{29–32} and free base (**Pheo a**)^{33,34} analogues of chlorophyll *a*, as well as benchmark zinc or free base chlorins (**Zn-8**, **8**)¹⁸ lacking a 13-substituent (Chart 1). The absorption spectra of the synthetic oxophorbines and chlorins are shown in Figure 1. The major observations are as follows:

(1) Absorption spectral effects of the 13-acetyl substituent: The presence of the 13-acetyl substituent in the zinc complex (**Zn-1** vs. **Zn-8**) red-shifts both the B band (12 nm) and the

CHART 1



Q_y band (27 nm), increases the intensity of the Q_y band (1.6-fold), and alters the I_B/I_{Q_y} ratio from 4.9 to 2.8. Similar trends are observed in the free base chlorins.

(2) Absorption spectral effects of the isocyclic ring: The presence of the isocyclic ring in the zinc complex (**Zn-2** vs **Zn-8**) red-shifts the Q_y band by 35 nm and increases the intensity of the Q_y band (1.5-fold). A similar trend was observed for the free base oxophorbine **2**, although the magnitude was less pronounced than for the zinc complexes given that the Q_y band of free base chlorin **8** appears at 641 nm. The absorption spectrum of the free base oxophorbine **2** closely resembles that of pheophorbide *a* (667 nm),^{33,34} a free base analogue of chlorophyll *a* (see Supporting Information for overlaid spectra).

(3) Fluorescence yields: The fluorescence quantum yield (Φ_f) increases by >4-fold in going from the benchmark zinc chlorin (**Zn-8**, $\Phi_f = 0.062$) to the zinc 13-acetylchlorin **Zn-1** or zinc oxophorbine **Zn-2**. The yields are comparable to that of the chlorophyll derivative **Zn-Pheo a** (0.23). The effect of introducing a keto substituent in the free base compounds is less pronounced, where the parent chlorin **8** is already rather emissive ($\Phi_f = 0.23$).

(4) IR properties: 13-Acetylchlorin **1** exhibits a carbonyl stretch (ν_{max}) at 1728 cm⁻¹, whereas that of oxophorbine **2** appears at 1701 cm⁻¹. The latter matches well with that of

(29) Boucher, L. J.; Katz, J. J. *J. Am. Chem. Soc.* **1967**, *89*, 4703–4708.

(30) Jones, I. D.; White, R. C.; Gibbs, E.; Denard, C. D. *J. Agric. Food Chem.* **1968**, *16*, 80–83.

(31) Dvornikov, S. S.; Knyukshto, V. N.; Solovov, K. N.; Tsvirko, M. P. *Opt. Spectrosc. USSR* **1979**, *46*, 385–388.

(32) Agostiano, A.; Catucci, L.; Colafemmina, G.; Scheer, H. *J. Phys. Chem. B* **2002**, *106*, 1446–1454.

(33) Eichwurz, I.; Stiel, H.; Röder, B. *J. Photochem. Photobiol. B: Biol.* **2000**, *54*, 194–200.

(34) Smith, J. H. C.; Benitez, A. In *Modern Methods of Plant Analysis*; Paech, K., Tracey, M. V., Eds.; Springer-Verlag: Berlin, 1955; Vol. IV, pp 142–196.

(25) (a) Muratake, H.; Natsume, M. *Tetrahedron Lett.* **1997**, *38*, 7581–7582. (b) Muratake, H.; Natsume, M.; Nakai, H. *Tetrahedron* **2004**, *60*, 11783–11803.

(26) Taniguchi, M.; Kim, M. N.; Ra, D.; Lindsey, J. S. *J. Org. Chem.* **2005**, *70*, 275–285.

(27) Weber, G.; Teale, F. W. J. *Trans. Faraday Soc.* **1957**, *53*, 646–655.

(28) Seely, G. R.; Jensen, R. G. *Spectrochim. Acta* **1965**, *21*, 1835–1845.

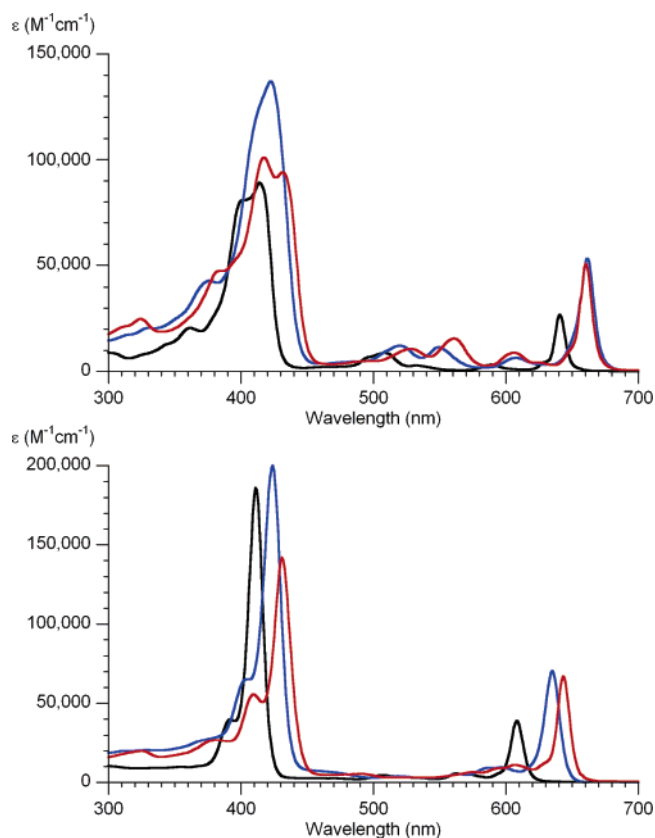


FIGURE 1. Absorption spectra in toluene at room temperature. Upper panel: free base compounds (color in graph; Q_y band position) include **8** (black, 641 nm); **1** (blue, 661 nm); **2** (red, 660 nm). Lower panel: zinc chelates include **Zn-8** (black, 608 nm); **Zn-1** (blue, 635 nm); **Zn-2** (red, 643 nm).

pheophytin *a* (1705 cm⁻¹),³⁵ methyl pheophorbide *a* (1703 cm⁻¹),³⁵ and methyl pyropheophorbide *a* (1695 cm⁻¹).³⁵

In summary, a new route has been developed for installing the isocyclic ring on tetrapyrrole macrocycles. The route entails preparation of a 13-acetylchlorin, which undergoes bromination at the 15-position followed by a Pd-mediated α -arylation procedure to close the ortho-perifused ring. The presence of a keto group at the 13-position significantly red-shifts the absorption maximum and increases the intensity of the Q_y band. The ability to install the isocyclic ring opens a number of possible applications ranging from artificial photosynthesis to photomedicine.

Experimental Section

13-Acetylation: 13-Acetyl-17,18-dihydro-10-mesityl-18,18-dimethyl-5-*p*-tolylporphyrin (1). Following a procedure for Stille coupling on aromatic compounds,²⁴ a mixture of **6** (19.8 mg, 0.0315 mmol), tributyl(1-ethoxyvinyl)tin (21.3 μ L, 0.0630 mmol), and (PPh₃)₂PdCl₂ (3.40 mg, 0.00472 mmol) was refluxed in THF (1.5 mL) for 20 h in a Schlenk line. The reaction mixture was treated with 10% aqueous HCl (1 mL) at room temperature for 2 h. CH₂Cl₂ was added, and the organic layer was separated. The organic layer was washed with saturated aqueous NaHCO₃, water, and brine. The organic layer was dried (Na₂SO₄), concentrated, and chromatographed [silica, hexanes then CH₂Cl₂/hexanes (1:1)], affording a purple solid (13.7 mg, 74%): IR 1728 cm⁻¹; ¹H NMR δ -0.98

(br, 2H), 1.86 (s, 6H), 2.02 (s, 6H), 2.61 (s, 3H), 2.66 (s, 3H), 3.05 (s, 3H), 4.56 (s, 2H), 7.24 (s, 2H), 7.50 (d, *J* = 7.8 Hz, 2H), 7.96 (d, *J* = 7.8 Hz, 2H), 8.23 (d, *J* = 4.4 Hz, 1H), 8.31 (d, *J* = 4.4 Hz, 1H), 8.68 (d, *J* = 4.4 Hz, 1H), 8.69 (s, 1H), 8.70 (d, *J* = 4.4 Hz, 1H), 8.86 (s, 1H), 9.98 (s, 1H); LD-MS obsd 590.6; FAB-MS obsd 590.3052, calcd 590.3046 (C₄₀H₃₈N₄O); λ_{abs} 423 (log ϵ = 5.14), 661 (4.72) nm; λ_{em} 665 (Φ_{f} = 0.25).

15-Bromination: 13-Acetyl-15-bromo-17,18-dihydro-10-mesityl-18,18-dimethyl-5-*p*-tolylporphyrin (7). Following a reported procedure,²⁶ a solution of **1** (26.4 mg, 0.0447 mmol) in THF (22 mL) was treated with NBS (7.95 mg, 0.0447 mmol) at room temperature for 1 h. CH₂Cl₂ was added. The mixture was washed with aqueous NaHCO₃. The organic layer was dried (Na₂SO₄), concentrated, and chromatographed [silica, hexanes then CH₂Cl₂/hexanes (3:1)], affording a purple solid (20.8 mg, 70%): ¹H NMR δ -1.30 to -1.40 (br, 1H), -0.98 to -1.02 (br, 1H), 1.86 (s, 6H), 2.04 (s, 6H), 2.60 (s, 3H), 2.66 (s, 3H), 3.06 (s, 3H), 4.55 (s, 2H), 7.21 (s, 2H), 7.51 (d, *J* = 7.8 Hz, 2H), 7.96 (d, *J* = 7.8 Hz, 2H), 8.26 (d, *J* = 4.4 Hz, 1H), 8.34 (d, *J* = 4.4 Hz, 1H), 8.45 (s, 1H), 8.70-8.76 (m, 3H); LD-MS obsd 668.4; FAB-MS obsd 668.2145, calcd 668.2150 (C₄₀H₃₇BrN₄O); λ_{abs} 418, 652 nm.

Installation of the Isocyclic Ring: 18,18-Dimethyl-10-mesityl-13¹-oxo-5-*p*-tolylporphorbine (2). A mixture of **7** (20.2 mg, 0.0301 mmol), Cs₂CO₃ (49.0 mg, 0.150 mmol), and (PPh₃)₂PdCl₂ (4.22 mg, 0.0125 mmol) was refluxed in toluene (3.0 mL) for 24 h in a Schlenk line. CH₂Cl₂ was added. The reaction mixture was washed with water and brine. The organic layer was dried (Na₂SO₄), concentrated, and chromatographed [silica, hexanes then hexanes/Et₂O (1:1)], affording a purple solid (15.1 mg, 85%): IR 1701 cm⁻¹; ¹H NMR δ -1.25 (br, 1H), 0.73 (br, 1H), 1.88 (s, 6H), 2.02 (s, 6H), 2.57 (s, 3H), 2.65 (s, 3H), 4.27 (s, 2H), 5.12 (s, 2H), 7.20 (s, 2H), 7.49 (d, *J* = 7.8 Hz, 2H), 7.93 (d, *J* = 7.8 Hz, 2H), 8.22 (d, *J* = 4.4 Hz, 1H), 8.28 (d, *J* = 4.4 Hz, 1H), 8.53 (s, 1H), 8.58 (s, 1H), 8.62 (d, *J* = 4.4 Hz, 1H), 8.70 (d, *J* = 4.4 Hz, 1H); LD-MS obsd 589.2; FAB-MS obsd 589.2971, calcd 589.2967 [(M+H)⁺, M = C₄₀H₃₆N₄O]; λ_{abs} 417 (log ϵ = 5.00), 431 (sh, 4.97), 660 (4.71) nm; λ_{em} 663 (Φ_{f} = 0.37).

Zinc Insertion: Zn(II)-10-Mesityl-18,18-dimethyl-13¹-oxo-5-*p*-tolylporphorbine (Zn-2). A solution of Zn(OAc)₂·2H₂O (106 mg, 0.484 mmol) in methanol (1.4 mL) was added to a solution of **2** (19.0 mg, 0.0323 mmol) in CHCl₃ (5.6 mL) with stirring at room temperature. After 16 h, the reaction mixture was concentrated, and CH₂Cl₂ was added. The organic layer was washed (saturated aqueous NaHCO₃, H₂O), dried (Na₂SO₄), and filtered. The filtrate was concentrated to dryness. The resulting solid was washed with hexanes several times. The solid was dissolved in a minimum amount of methanol, and then hexanes was added (~2:1 ratio of hexanes/methanol), affording a green solid (18.0 mg, 85%): ¹H NMR (THF-*d*₈) δ 1.88 (s, 6H), 2.03 (s, 6H), 2.56 (s, 3H), 2.63 (s, 3H), 4.30 (s, 2H), 5.00 (s, 2H), 7.22 (s, 2H), 7.47 (d, *J* = 7.6 Hz, 2H), 7.88 (d, *J* = 7.6 Hz, 2H), 8.15 (d, *J* = 4.4 Hz, 1H), 8.18 (d, *J* = 4.4 Hz, 1H), 8.33 (s, 1H), 8.48 (s, 1H), 8.50 (d, *J* = 4.4 Hz, 1H), 8.62 (d, *J* = 4.4 Hz, 1H); LD-MS obsd 650.2; FAB-MS obsd 650.2031, calcd 650.2024 (C₄₀H₃₄N₄OZn); λ_{abs} 409 (sh, log ϵ = 4.73), 431 (5.15), 643 (4.84) nm; λ_{em} 646 (Φ_{f} = 0.28).

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Supporting Information Available: Complete experimental procedures; description of porphyrin nomenclature; crystallographic data for **Cu-1**. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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(35) Katz, J. J.; Dougherty, R. C.; Boucher, L. J. In *The Chlorophylls*; Vernon, L. P., Seely, G. R., Eds.; Academic Press: New York, 1966; pp 185-251.